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Determination of Grouper Species of Subfamily Epinephelinae from Raja Ampat (West Papua) Region Using CO1 Gene Sequence

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Abstract

The Raja Ampat Islands, located near the heart of the "Coral Triangle" on Bird's Head Peninsula, West Papua, Indonesia are well known for its outstanding biological diversity and stunning marine and terrestrial habitats. Groupers (family Serranidae) has included as part of the five largest families associated with coral reefs on the Bird's Head region. The grouper identification was generally made based on color patterns and morphological characters, but often, these characters show intraspecific variations or differences in color patterns between juvenile and adult individuals. This study aims to confirm the type of grouper species obtained from the sport fishing activities around Raja Ampat Island. Species confirmation from the subfamily of Epinephelinae determined by analyzing the number of differences of nucleotides and genetic distance on the sequence of the CO1 gene (Cytochrome oxidase subunit 1). A total of eight fish samples were successfully sequenced and aligned. Those samples consist of eight species belonging to the three genera, namely Anyperodon, Epinephelus, and Cephalopolis.

Key Words: Epinephelinae, CO1, sports fishing, genetic distance, grouper, nucleotide

Introduction

The Raja Ampat Islands, located near the heart of the "Coral Triangle" on Bird's Head Peninsula, West Papua, Indonesia are well known for its outstanding biological diversity and stunning marine and terrestrial habitats. The Raja Ampat Island composed of four main islands (Waigeo, Batanta, Salawati, and Misool) and hundreds of smaller islands. The archipelago has one of the highest diversity of the world's coral reef fish faunas, which consisted of at least 1074 species. Groupers (family Serranidae), including into the five largest families associated with coral reefs in the Bird's Head region (McKenna *et al.* 2002).

The Serranid are a large family of fishes belonging to the order Perciformes. The Serranid Subfamily Epinephelinae commonly known as groupers, rockcods, hinds, and sea basses, comprises about 159 species of marine fishes in 15 genera. These commercially essential fishes are bottom-associated, which found in tropical and subtropical waters. Most species occupy coral reefs, but some inhabit estuaries or on rocky reefs (Heemstra and Randall 1993). Grouper has potential economic value in fisheries, many species such as Epinephelus coioides, E. E. malabaricus, tauvina, E.marginatus, Cephalopholis boenak known as consumption fish while the others also used as ornamental fish (Hemstra and Randal 1993; Asensio et al. 2009; Noitokr et al. 2013). Color patterns and morphological characters generally used for the grouper identification, but overlapping meristic counts and changes in a color pattern during life stages contributes to misidentification in grouper species (Hemstra & Randall 1993, Craig *et al.* 2001, Alcantara & Yambot 2014). It causes any kind fishes of this subfamily member caught in the field are often summed up as groupers (Ariyanti 2015)

A partial sequence of the mitochondrial cytochrome oxidase c subunit 1 (CO1) gene commonly used as a barcode with the size is about 650 bp. CO1 gene has widely used in several animal taxa such as insects, birds, and fish (Hebert *et al.* 2007). This gene has also been used in rapid analyses for commercial purposes, especially for the confirmation of fish species (Ward *et al.* 2005; Barber and Boyce 2006; Wong and Hanner 2008; Sachithanandam *et al.* 2012). This present work applied the partial CO1 as a molecular marker to delimitate the type of grouper species that obtained from sport fishing activities in around Raja Ampat Island.

Materials and Method

Fish species belonging to family Serranidae were collected from several sampling points in Raja Ampat areas specifically around Salawati island (Fig.1). Phenotypic characterization was analyzed using FAO species catalog of groupers of the world. Tissue samples that were used as the source of DNA are part of the dorsal muscle tissue. All tissue specimen was stored within 95% alcohol for the molecular genetic study.



Figure 1. Location of Salawati Island in Bird's Head Peninsula, West Papua, Indonesia.

DNA Extraction and PCR Reaction

Total DNA was extracted from 0.30 g of ethanol-preserved tissue muscle using DNA Extraction Kit for animal tissue (Geneaid) by following the manufacturer's protocol. Approximately 650-655 bp were amplified from the 5' region of the COI gene using combinations of the fish-specific primers AF282 and AF283 which is designed as a degenerate primer for fish were described in Ward *et al.* (2005).

The 50 μ L PCR mixes included 25 μ L 2X GoGreen GoTaq DNA polymerase mix (0.05 U/ μ L, 3mM Mg²⁺, 0.4 mM each dNTP), 1 μ L of each primer (1st BASE, Singapore), 2 μ L of DNA template and 21 μ L of Nuclease-Free Water. The thermal regime consisted of an initial step of 2 min at 94 °C followed by 36 cycles of 0.5 min at 94 °C, 0.5 min at 56-57 °C, and 1 min at 72 °C, followed in turn by 7 min at 72 °C and then held at 4.0 °C in an Applied Biosystems (Foster City, CA, USA) EscoTM thermocycler.

Visualization

The amplicon was performed using a 6% polyacrylamide gel that runs at a voltage of 200 volts for 40 minutes. Afterward, proceed with silver staining.

Data Analysis

All amplicon were sequenced through company service sequencing following the manufacturer's protocol. DNA sequences were proofread, aligned, and edited using BioEdit (Hall 1999). Multiple alignments were done using Clustal W in MEGA 6 (Tamura *et al.* 2013). Sequence divergence was estimated using the

Kimura two Parameters model of base substitution (Kimura, 1980). Phylogenetic tree reconstruction was done using the distance-based method, Neighbor-joining (NJ), and Maximum Likelihood (ML) as comparative trees base on CO1 region with nodes frequencies were calculated based on 1000 bootstrap replicates. Several sequences of grouper species used in GenBank (accession number in the figure) to root the tree for comparative purposes.

Result and Discussion

Each sequence in this work has aligned with an accuracy of homologous analysis result sequences from the GenBank (BLAST) about 98-100%. Those samples confirmed as Epinephelus coeruleopunctatus (RJ1), E. ongus (RJ5), E. fasciatus (RJ6), E. melanostigma (RJ7), E. quoyanus (RJ9), E. coioides (RJ4), Anyperodon leucogramicus (RJ3), and Cephalopholis miniata (RJ11). The partial CO1 sequences were 520 nucleotides (nt) with 188 nt variable sites, 332 nt sites. 169 nt parsimony conserved and informative sites. Based on the partial CO1 and using Haemulon scuderii as an outgroup, a molecular phylogenetic tree constructed by the Neighbor-Joining (NJ) method (Kimura 2values of the bootstrap parameter). The confidence level of nodes indicated above the branch. Fig. 2 shows that all GenBank sequences and sequences of subfamily Epinephelinae acquired in this study. NJ tree exhibited 2 clades that separate 3 genera (including Anyperodon).

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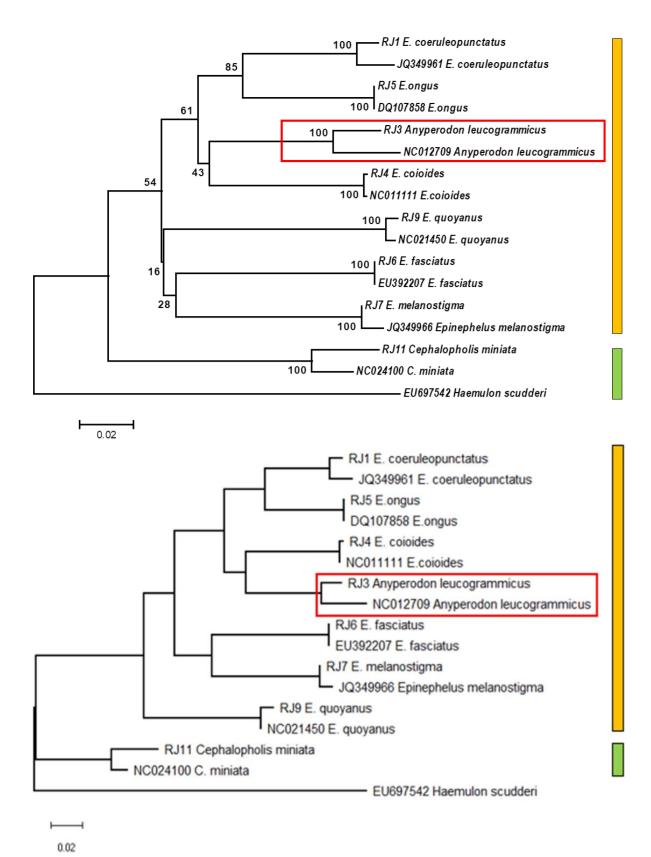


Figure 2. Neighbor-joining and Maximum Likelihood (ML) as comparative trees based on the mtDNA CO1 nucleotide sequences of groupers samples. Numbers at nodes are bootstrap values on 1000 replicates.

Table 1. Number of differences of nucleotides (below the diagonal) and genetic distances of CO1 (above the diagonal) in present work

No	Species	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17
1	RJ1 (E. coeruleopunctatus)		0.139	0.127	0.099	0.175	0.153	0.165	0.170	0.151	0.175	0.099	0.162	0.157	0.125	0.022	0.183	0.262
2	RJ3 (Anyperodon leucogrammicus)	64		0.117	0.135	0.159	0.169	0.167	0.205	0.042	0.159	0.135	0.179	0.164	0.119	0.148	0.184	0.245
3	RJ4 (E. coioides)	59	55		0.119	0.146	0.152	0.167	0.217	0.131	0.146	0.119	0.161	0.165	0.002	0.134	0.201	0.265
4	RJ5 (<i>E.ongus</i>)	47	63	56		0.174	0.127	0.175	0.204	0.143	0.174	0.000	0.136	0.172	0.121	0.105	0.204	0.265
5	RJ6 (E. fasciatus)	79	73	68	79		0.142	0.159	0.212	0.164	0.000	0.174	0.151	0.162	0.144	0.174	0.191	0.254
6	RJ7 (E. melanostigma)	70	77	70	60	66		0.159	0.206	0.169	0.142	0.127	0.008	0.167	0.150	0.159	0.209	0.236
7	RJ9 (E. quoyanus)	75	76	76	79	73	72		0.181	0.169	0.159	0.175	0.168	0.008	0.165	0.172	0.168	0.301
8	RJ11 (Cephalopholis miniata)	78	91	96	91	94	91	82		0.208	0.212	0.204	0.216	0.178	0.215	0.183	0.040	0.266
9	NC012709 Anyperodon leucogrammicus	69	21	61	66	75	77	77	92		0.164	0.143	0.179	0.167	0.134	0.156	0.184	0.262
10	EU392207 E. fasciatus	79	73	68	79	0	66	73	94	75		0.174	0.151	0.162	0.144	0.174	0.191	0.254
11	DQ107858 E.ongus	47	63	56	0	79	60	79	91	66	79		0.136	0.172	0.121	0.105	0.204	0.265
12	JQ349966 Epinephelus melanostigma	74	81	74	64	70	4	76	95	81	70	64		0.176	0.159	0.159	0.219	0.247
13	NC021450 E. quoyanus	72	75	75	78	74	75	4	81	76	74	78	79		0.162	0.164	0.165	0.298
14	NC011111 E.coioides	58	56	1	57	67	69	75	95	62	67	57	73	74		0.131	0.198	0.267
15	JQ349961 E. coeruleopunctatus	11	68	62	50	79	73	78	83	71	79	50	73	75	61		0.181	0.274
16	NC024100 C. miniata	83	83	90	91	86	92	77	20	83	86	91	96	76	89	82		0.251
17	EU697542_Haemulon_scudderi	114	108	115	115	111	105	127	115	114	111	115	109	126	116	118	110	

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Based on table 1, the smallest number of differences nucleotides was 47 nt, between RJ1 and RJ5. Then the biggest is 96 nt between RJ4 and RJ11. The differences number of nucleotides seem like appropriate with genetic distances. Minimum genetic distances value in CO1 among all *Epinephelus* genera was 0.099 between RJ1 (*E. coeruleopunctatus*) and RJ5 (*E. ongus*). The maximum genetic distances value is RJ5 and RJ9, RJ1 and RJ6 with genetic distance 0.175 respectively. The maximum pairwise nucleotide divergence value between RJ4 and RJ11 is 0.217.

Neighbor-Joining tree exhibited 2 clades that separate 3 genera (including Anyperodon). The phylogenetic analysis showed the lowest genetic distance between Epinephelus and Anyperodon (Table 1). It shows in Fig. 2 that Anyperodon leucogrammicus was grouped within Epinephelus genera so that the cluster become not monophyletic. This fact confirms the paraphyletic status of the Epinephelus (Craig et al. 2001; Zhu et al. 2008; You et al. 2013). Current classification, Anyperodon is distinctive monotypic genus is probably most closely related to Epinephelus, with which it shares XI dorsal-fin spines and the absence of trisegmental pterygiophore, but it differs from Epinephelus (and all other groupers) in its missing teeth on the palatines. Anyperodon is also unique among groupers in its elongate groupers, but none of these specimens are looking as compressed as Anyperodon (Hemstra & Randall 1993).

The position Epinephelus and Cephalopholis in the phylogenetic tree also similar with Craig et al. (2001), using 16S gene and then confirmed by Craig and Hasting (2007) that support the valid genus of the Cephalopholis separate from Epinephelus. Cephalopholis is primitive than genus Epinephelus. Cephalopholis is one of the wealthiest genera (besides Mycteroperca and Epinephelus) which has various species. The NJ tree shows the C. miniata become sister clades of Epinephelus clades. The position of Epinephelus at the top of the phylogenetic tree indicating that is the most

References

- Alcantara SG, Yambot AV. 2016. Dna barcoding of commercialy important Grouper species (Percifromes, Serranidae) in the Philippines. Mitochondrial DNA 27 (6): 3837-3845.
- Ariyanti Y. 2015. Phylogenetic analysis of some Indo West-Pacific grouper subfamily Epinephelinae (Serranidae) from Indonesian waters. Animal Biosciences, Graduate School, Bogor Agricultural University.

recently diverged species, which is in concordant with the fact that it is also the most advanced genus in Epinephelinae (Craig *et al.* 2001; Craig and Hasting 2007, Ding *et al.* 2006).

E. quoyanus is one of 9 shallow-water coral reef species that have a rounded caudal fin and close-set dark brown spots with the pale interspaces forming a network on the body. These reticulated groupers have been much confused in the literature, and many museum specimens have misidentified with the other species such as E. bilobatus, E. faveatus, E. hexagonatus, E. macrospilos, E. maculatus, E. melanostigma, E. merra, and E. spilotoceps (Hemstra and Randall 1993).

The difficulties of groupers species identification and confirmation based morphological characters could be resolved by identification using partial CO1. Also, the genetic relationship of the groupers successfully reconstructed through the phylogenetic analysis. Partial cytochrome oxidase c subunit 1 (CO1) gene successfully delimitate the type of grouper species and reveal the diversity of groupers species that obtained from the sport fishing activities in around Raja Ampat Island.

Conclusion

Species confirmation and determination of subfamily Epinephelinae did by analyzing the differences between nucleotide and genetic distance on the segment of the CO1 gene (Cytochrome oxidase subunit 1). A total of eight fish samples were successfully sequenced and aligned with data from the GenBank. Those samples consist of eight species belonging to three genera, namely *Anyperodon*, *Epinephelus*, and *Cephalopolis*. This work exhibit the exceptional diversity of groupers species from Raja Ampat, as being shown from the samples acquired from sport fishing activities.

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- Asensio L, Gonzalez I, Rojas M, Garcia T, Martin R. 2009. PCR-based methodology for the authentication of grouper (*Epinephelus marginatus*) in commercial fish fillets. Food cont 20 (7): 618-622.
- Barber P, Boyce SL. 2006. Estimating diversity of Indo-Pacific coral reef stomatopods through DNA barcoding of stomatopod larvae. Proc R Soc Lond B. 273: 2053-2061.
- Craig MT, Pondella DJ, Franck JPC, Hafner JC. 2001. On the Status of the Serranid Fish Genus *Epinephelus*: Evidence for

- paraphyly Based upon 16S rDNA sequence. *Mol Phyl Evol.* 19: 121-130.
- Craig MT, Hastings PA. 2007. Molecular phylogeny of the groupers of the subfamily Epinephelinae (Serranidae) with a revised classification of the Epinephelini. *Ichthyol Res.* 54: 1-17.
- Ding SX, Wang YH, Wang J, Zhuang X, Su YQ, You YZ, Li QF. 2006. Molecular phylogenetic relationships of 30 gruper species in China Seas based on 16S rDNA fragment sequences. *Acta Zoologi Sinica*.52 (3): 504-513.
- Hall TA. 1999. Bioedit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. *Nucl Acids Symp Ser.* 4: 95-98.
- Heemstra PC, Randall JE. 1993. FAO species catalogue: An annotated and illustrated catalogue of the grouper, rockcod, hind, coral grouper, and lyretail species known to date. Volume 16 Groupers of the world (Family Serranidae, subfamily Epinephelinae). Rome (IT): FAO of the United Nations.
- Kimura M. 1980. A simple method for estimating evolutionary rate of base edsubstitions trough comparative studies of nucleotide sequences. J Mol Evol. 16: 111-120.
- McKenna SA, GR Allen and S Suryadi (eds). 2002. A Marine Rapid Assessment of the Raja Ampat Islands, Papua Province,

- Indonesia. RAP Bulletin of Biological Assessment 22. Conservation International, Washington, DC.
- Sachithanandam V, Mohan PM, Chaaithanya IK, Dhivya P, Baskara R. 2012. DNA barcoding, phylogenetic study of *Epinephelus* spp. from Andaman coastal region, India. *Indian J Geo Mar Sci.* 41(3): 203-211.
- Tamura K., Stecher G., Peterson D., Filipski A., & Kumar S. 2013. MEGA6: Molecular Evolutionary Genetics Analysis version 6.0. *Molecular Biology and Evolution*30: 2725-2729.
- Ward RD, Zemlak TS, Innes BH, Last PR, Hebert PD. 2005. DNA Barcoding of Australia's fish species. *Phil Trans R Soc B.* 360: 1847-1857.
- Wong EHK, Hanner RH. 2008. DNA barcoding detect market substitution in North American seafood. *Food Res Int.* 41: 828-837.
- You X, Shu L, Li S, Chen j, Lou J, Lu J, Mu Q, Bai J, Xia Q, Chen Q. 2013. Construction of high-density genetic linkage maps for orange-spotted grouper *Epinephelus coioides* using multiplex shotgun genotyping. *BMC Genetics*. 14: 113.
- Zhu S, Yang Y, Zheng W, Shen X, Zou J. 2008. Molecular phylogenetic relationship of *Epinephelus* base on sequences of mtDNA Cyt b. *Front Biol China*. 3 (2): 207-2.